

# Effect of Cooking Time on Biochemical Parameters and Some Functional Properties of Lima Bean Seed Flours (White, Red and Black) Consumed in Côte d'Ivoire

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**Abstract:** Meeting the world's food needs requires efforts to promote the cultivation and use of underutilized and neglected plants, which have the potential to improve food and nutritional security. *Phaseolus lunatus* (L.), is one of them can serve as a promising potential food crop. This study aims to contribute to food security through the valorization of *P. lunatus* (L.) beans seeds, with a view to their more rational use in different diets. Seeds obtained at stage 4 (52 days) of maturity after harvesting were cooked for 45, 60 and 75 minutes at 100°C. The results showed an increase in moisture content (29.68 to 36.27%), carbohydrates (64.16 to 71.56%) and fiber (4.62 to 6.05%), followed by a reduction in protein (4.62 to 6.05%), ash (4.62 to 6.05%) and lipids (4.62 to 6.05%). The results also showed a non-significant decrease in essential and non-essential amino acids during cooking. Similarly, cooking increased water absorption capacity (WAC) (230.00 to 322.60%), oil absorption capacity (OAC) (190.36 to 250.36%), swelling power (SP) (2.26 to 23.33 g water/g DM) and solubility (1.80 to 19.90%) at temperatures ranging from 50 to 90°C. The study provides useful information for consumers and food manufacturers that lima bean flour has great potential to increase the nutritional value of foods.

**Keywords:** Cooking, Neglected Plants, Underutilized, Food Security, Food Manufacturers

## 1. Introduction

According to [1], dry beans are one of the main legumes that are an essential source of nutrients and dietary protein for over half a billion people worldwide [2]. Dried bean seeds are good source of energy, complex carbohydrates (dietary fiber, starch and oligosaccharides), proteins, minerals and vitamins as well as polyphenols and antioxidants necessary for human health [3]. *Phaseolus lunatus* is a species of dry bean, belonging to the Fabaceae family and the *Phaseolus* genus. Its seeds are an important source of protein (21-26%), carbohydrates (55-64%), fiber (3.2-6.8%), lipids (12.3%) and minerals such as potassium, zinc, calcium, iron with small amounts of sodium and phosphorus [4]. They are used in many preparations as food

supplements with cereals such as rice, maize, sorghum or with plantain. *P. lunatus* is grown in association with tubers, plantain and vegetables in the forest zone, but also with sorghum and millet in the savannah zone [5]. High consumption of *P. lunatus* seeds reduces the risk of developing diabetes, hypertension and hypercholesterolemia, according to [6]. They are also particularly rich in essential amino acids such as lysine [7]. Various mechanisms involved in food preparation can affect the nutritional value and bioavailability of nutrients and minerals. Previous studies have shown that technological processes such as soaking and cooking have an impact on the nutritional value of foods. The aim of this study was to evaluate the effects of different cooking times on the biochemical parameters and some functional properties of *P. lunatus*.

## 2. Material and Methods

### 2.1. Materials

Seeds of the three *Phaseolus lunatus* (L.) cultivars (white,

red and black) were obtained from pods harvested at full withering in a field created in Tomasset, 38.7 km from Abidjan, during 2013-2014 [8].

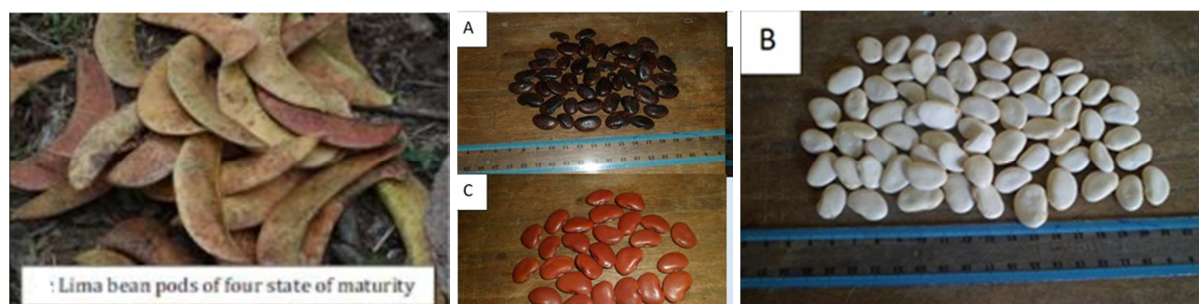


Figure 1. Lima bean pods of four state of maturity, A=black seeds, B=white seeds and C=red seeds.

### 2.2. Methods

#### 2.2.1. Preparation of Lima Bean Powder

Three hundred (300) grams of each sample of, lima bean (*Phaseolus lunatus*) white, red and black bean seeds at maturity stage 4 (52 days) were mixed in an ivory pot with 2 liters of pre-boiled water (100°C) Cook together. On a hot plate (RELPE, Spain). Precook to determine different cooking times. The 45minutes time was marked by removing the seed membrane by rubbing with two fingers. Fifteen (15) minutes are considered the cooking interval. Therefore, three cooking times (45, 60, 75 minutes) were determined. Put the seeds in the IVOIRAL, pot. As soon as cooking starts, the timer starts. After each cooking time (45, 60 and 75 minutes), remove the seeds and drain for a few minutes. After draining, the seeds were dried in a ventilated oven at 45°C for 72 h. They are ground with a Moulinex grinder. The powder obtained was sieved with an AFNOR 300 µm sieve. The flour obtained was stored in glass bottles for laboratory analysis, previously washed and dried in an oven at 45°C [8].

#### 2.2.2. Proximate Analysis of Samples

Moisture, ash, crude protein, crude fat, crude fiber and total sugar were determined respectively by standard method [9,10], and carbohydrate content was determined by difference equation  $([100 - (\text{protein} + \text{crude fat} + \text{ash} + \text{crude fiber})])$  [11]. In addition, energy value (EV) was calculated by applying the calorie coefficient from [12] according to the following equation:  $\text{EV (Kcal/100g)} = (4 \times \text{protein}\%) + (4 \times \text{carbohydrate}\%) + (9 \times \text{fat}\%)$ . Analyses values are triplicate values.

#### 2.2.3. Mineral Analysis

Minerals were analyzed using the method described in [10]. Take 1 g of ash obtained from the sample, dissolve it in 10% HCl, filter it with filter paper and delute to the standard volume with deionized water. The content of sodium and potassium in the samples were determined by flame photometry described method in litterature [13]. Calcium, Iron, Magnesium, Zinc and Cooper were determined using Atomic Absorption Spectrophotometry (AAS). Phosphorus content was estimated colorimetrically using spectrophotometry (UV-visible

spectrophotometer, Mode IDR 2800/United States).

#### 2.2.4. Amino Acid Analysis

The amino acid contents of the samples were determined using an automatic amino acid analyzer (BIOCHROM30, serial number103274), according to the method described in literature [14].

#### 2.2.5. Functional Properties Evaluation

##### (i). Water Absorption Capacity (WAC) and Water Solubility Index (WSI)

The water absorption capacity (WAC) and water solubility index (WSI) of flour were determined according to [15] using a simple (modified) technique. Dissolve one gram (1 g) of flour (M0) in 10 mL of distilled water in a centrifuge tube. The mixture was stirred with a shaker for 30 minutes, and then kept in a water bath at 37°C for 30 minutes. Then, centrifuge at 3000 rpm for 15 minutes with a centrifuge (ORTOAL RESAR). Weigh the resulting pellet (M2) and then dry it in a 105°C oven for 24 h until a constant mass (M1) is reached. CAE and ISE are calculated according to the following relationship:

$$\text{CAE (\%)} = \frac{(M2 - M1)}{M1} \times 100$$

$$\text{ISE (\%)} = \frac{(M0 - M1)}{M0} \times 100$$

##### (ii). Oil Absorption Capacity

The oil absorption capacity (OAC) of lima bean flours was evaluated using the method of [16] methods. Mix 1g of sample (M0) with 10 ml of oil. The slurry was stirred on a vortex mixer for 2 minutes, allowed to stand at room temperature (28°C) for 30 minutes and then centrifuged at 15,000 rpm for 10 minutes. Weighed the sediment (M1). The oil holding capacity is calculated according to the following formula:

$$\text{CAH (\%)} = \frac{(M1 - M0)}{M0} \times 100$$

##### (iii). Swelling and Solubility

Swelling and solubility tests were performed according to the technique of [17] with slight modifications. A

suspension of 1% (w/v) flour was placed in a centrifuge tube and then agitated maximally for 30 min in a water bath at different temperatures (50, 60, 70, 80 and 90°C). Centrifuge the suspension in the tube at 3,000 rpm for 15 min. Aliquots of the pellet and supernatant were then collected in separate containers and the supernatant was incubated at 105°C for 24 hr and the pellet for 48 hr. The supernatant was used to determine solubility and swelling:

$$PG = \frac{(M1-M2)}{M1} \times 100$$

PG: swelling power (gwater/g starch);

M1: wet mass of aliquot (g)

M2: aliquot dry mass (g)

$$S (\%) = \frac{M_s}{M} \times 100$$

S: solubility expressed as a percentage (%)

Ms: sample mass after drying (g)

M: mass of sample taken (g) to prepare 1%(w/v) solution

### 2.3. Statistical Analysis

The analysis of variance (ANOVA) was used to determine the differences between treatments. When a difference was observed, the multiple range tests of Newman-Keuls at 5% were performed to separate treatment means. Statistical tests were performed using the STATISTICA software version 7.1.

## 3. Results

### 3.1. Effect of Cooking Time on the Biochemical Composition of Phaseolus lunatus (L.) Bean Seeds

The determination of biochemical components of *P. lunatus* bean seeds showed that with the prolongation of cooking time, the contents of total sugar, reducing sugar, protein, lipid and ash all decreased significantly ( $P < 0.05$ ) (Table 1). Total and reducing sugars in cooked *P. lunatus* (L) bean seeds decreased from  $4.46 \pm 0.75$  to  $2.90 \pm 0.23\%$  of dry matter and from  $0.53 \pm 0.75$  to  $0.30 \pm 0.13\%$ , respectively (Figures 1 and 2). Seed

protein content ranged from  $25.06 \pm 0.13$  to  $18.52 \pm 0.23$  mg/100 g dry matter. Protein losses from cooked seeds ranged from 10.22% to 10.27%. The lipid content of *P. lunatus* seeds ranged from  $2.16 \pm 0.08$  to  $0.69 \pm 0.01$  mg/100 g dry matter. The lipid lowering rate of seeds was between 49.07% and 50.71%. The ash content of *P. lunatus* seeds decreased from  $4.26 \pm 0.05$  to  $2.88 \pm 0.04$  mg/100 g dry matter. If the seeds were boiled for 45 minutes, there was minimal loss of protein, lipid and ash regardless of the seed. A cooking time of 45 minutes is adequate for seeds of good nutritional quality. Fiber, water and carbohydrate content increased significantly ( $P < 0.05$ ) during cooking. Dietary fiber values were  $4.62 \pm 0.04$  to  $6.05 \pm 0.13$  mg/100 g dry matter, moisture  $29.68 \pm 0.75$  to  $36.27 \pm 0.25$  mg/100 g dry matter, and  $64.16 \pm 0.09$  to  $71.56 \pm 0.04$  mg /100 g carbohydrate dry matter. Red seeds are rich in carbohydrates, ranging from  $69.63 \pm 0.48$  to  $71.56 \pm 0.04$  mg /100 g dry matter. When *P. lunatus* seeds were boiled for 75 minutes, the water, carbohydrate and fiber content increased. Cooking time had no effect on moisture, lipid or total carbohydrate content. On the other hand, energy values increased significantly ( $P < 0.05$ ) up to 45 min cooking time ( $300.79 \pm 0.59$  –  $374.28 \pm 0.27$  Kcal / 100 g dry matter), and then decreased ( $374.28 \pm 0.27$  –  $365.37 \pm 0.06$  mg/ 100 grams of dry matter) until the end of the cooking time. The high carbohydrate content of the red seeds gives them a high energy value with cooking time (45) minutes. With a cooking time of 45 minutes, the white seeds are very high in protein and fiber.

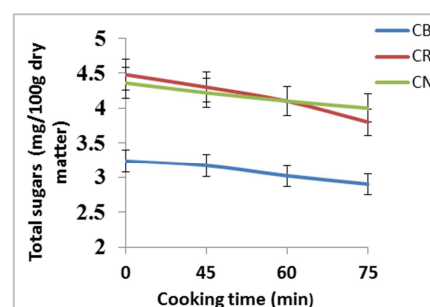


Figure 2. Effect of cooking time on total sugars (mg/100g dry matter) in *Phaseolus lunatus* bean seeds.

Table 1. Effect of cooking time on biochemical composition of *Phaseolus lunatus* bean seeds (g/100g dry matter).

Flours	Cooking time (min)	Moisture (%)	Proteins (%)	Fats (%)	Carbohydrates (%)	Fibers (%)	Ashs (%)	Value Energy (Kcal/100g)
FCW	0	29.68±0.75 <sup>d</sup>	25.06±0.13 <sup>a</sup>	1.40±0.01 <sup>c</sup>	64.16±0.09 <sup>b</sup>	5.13±0.07 <sup>de</sup>	4.17±0.00 <sup>b</sup>	300.79±0.59 <sup>e</sup>
	45	30.46±0.26 <sup>cd</sup>	23.60±0.10 <sup>b</sup>	1.03±0.06 <sup>de</sup>	65.63±0.25 <sup>a</sup>	5.91±0.08 <sup>fg</sup>	3.88±0.07 <sup>f</sup>	366.67±0.14 <sup>i</sup>
	60	31.20±0.20 <sup>bc</sup>	22.68±0.13 <sup>c</sup>	0.81±0.02 <sup>f</sup>	66.92±0.27 <sup>d</sup>	6.00±0.10 <sup>g</sup>	3.60±0.03 <sup>gh</sup>	365.60±0.20 <sup>j</sup>
	75	31.78±0.13 <sup>b</sup>	22.01±0.10 <sup>d</sup>	0.69±0.01 <sup>f</sup>	67.79±0.28 <sup>cd</sup>	6.05±0.13 <sup>g</sup>	3.42±0.07 <sup>ch</sup>	365.37±0.06 <sup>j</sup>
FCR	0	33.30±0.34 <sup>f</sup>	20.63±0.20 <sup>f</sup>	1.60±0.01 <sup>b</sup>	69.63±0.48 <sup>c</sup>	4.62±0.04 <sup>a</sup>	4.26±0.05 <sup>b</sup>	311.25±0.33 <sup>a</sup>
	45	33.90±0.26 <sup>ef</sup>	19.59±0.10 <sup>gh</sup>	1.08±0.10 <sup>c</sup>	70.37±0.12 <sup>cd</sup>	5.05±0.13 <sup>de</sup>	3.91±0.02 <sup>fg</sup>	369.60±0.40 <sup>d</sup>
	60	34.20±0.36 <sup>ef</sup>	18.90±0.03 <sup>ij</sup>	0.85±0.03 <sup>df</sup>	71.15±0.05 <sup>gh</sup>	5.50±0.20 <sup>bc</sup>	3.63±0.05 <sup>h</sup>	367.85±0.22 <sup>b</sup>
	75	34.63±0.21 <sup>e</sup>	18.52±0.23 <sup>j</sup>	0.70±0.03 <sup>f</sup>	71.56±0.04 <sup>g</sup>	5.83±0.15 <sup>fg</sup>	3.40±0.02 <sup>cd</sup>	366.96±0.31 <sup>hi</sup>
FCB	0	35.36±0.33 <sup>ge</sup>	21.21±0.18 <sup>e</sup>	2.16±0.08 <sup>a</sup>	67.95±0.28 <sup>f</sup>	4.93±0.08 <sup>ac</sup>	3.74±0.05 <sup>fg</sup>	310.02±0.68 <sup>f</sup>
	45	35.89±0.01 <sup>g</sup>	19.70±0.10 <sup>g</sup>	1.68±0.03 <sup>b</sup>	70.09±0.08 <sup>fg</sup>	5.32±0.07 <sup>cd</sup>	3.21±0.02 <sup>de</sup>	374.28±0.27 <sup>a</sup>
	60	36.10±0.34 <sup>g</sup>	19.22±0.06 <sup>ef</sup>	1.35±0.05 <sup>c</sup>	70.82±0.17 <sup>egh</sup>	5.60±0.08 <sup>bcd</sup>	3.00±0.10 <sup>e</sup>	372.31±0.40 <sup>b</sup>
	75	36.27±0.25 <sup>g</sup>	19.03±0.06 <sup>hi</sup>	1.10±0.10 <sup>c</sup>	71.12±0.11 <sup>gh</sup>	5.86±0.12 <sup>fg</sup>	2.88±0.04 <sup>a</sup>	370.59±0.52 <sup>c</sup>

Mean ± SD, n = 3; in columns, means marked with different letters indicate significant differences at the threshold ( $P < 0.05$ ). FCW (White Cultivar Flour), FCR (Red Cultivar Flour) and FCB (Black Cultivar Flour).

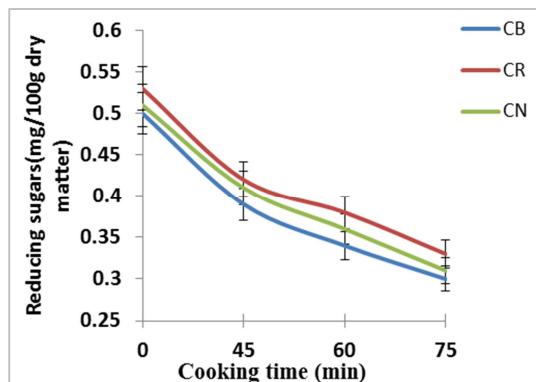


Figure 3. Effect of cooking time on reducing sugars (mg/100g dry matter) in *Phaseolus lunatus* bean seeds.

### 3.2. Effect of Cooking Time on Mineral Composition of *Phaseolus Lunatus* Bean Seeds

Analysis of seed meal cooked for 45, 60 and 75 minutes showed a decrease in potassium, phosphorus, magnesium, sodium, calcium and iron during cooking (Table 2). Potassium ( $1109.3 \pm 0.81$ - $1592.27 \pm 6.38$  mg/100 g dry matter); phosphorus ( $230.18 \pm 1.13$ - $259.25 \pm 1.08$  mg/100 g dry matter); calcium ( $301.03 \pm 1.00$  -  $359.32 \pm 0.89$  mg/100 g dry

matter); magnesium ( $128.46 \pm 0.35$  -  $140.60 \pm 0.52$  mg/100 g dry matter), sodium ( $60.80 \pm 0.60$  -  $75.27 \pm 0.72$  mg/100 g dry matter) and iron ( $9.49 \pm 0.27$  -  $12.56 \pm 0.28$  mg/100 g dry matter) of uncooked seeds (FNC) were significantly higher than cooked seeds ( $P < 0.05$ ). Mineral loss rates varied respectively from (34.71 - 46.48%) potassium, (42.05 - 46.53%) phosphorus, (35.46 - 42.31%) magnesium, (34.98 - 40%) calcium and iron (48.36- 57.40%). The mineral loss rates of cooked *P. lunatus* seeds are statistically different ( $P < 0.05$ ). At 45 minutes cooking time for seeds of the black cultivar, the sodium, potassium and magnesium losses obtained varied by 9.29%, 34.70% and 35.46% respectively. In contrast, cooking the seeds of the black cultivar for 45 minutes resulted in calcium, phosphorus and sodium losses of 34.98%, 42.05% and 11.60% respectively. Cooking the seeds caused more iron loss with white seeds (57.40%) than with black and red seeds. Na/K and Ca/P ratios vary by (0.05-0.06) and (1.31-1.41) respectively in uncooked seeds, while the same ratios vary by (0.05-0.07) and (1.21-1.63) respectively in cooked seeds. Minerals do not have the same sensitivity to heat. After 45 minutes of cooking, seeds from all three cultivars show high mineral concentrations.

Table 2. Effect of cooking time on mineral composition of *Phaseolus lunatus* bean seeds (mg/100g dry matter).

Flours	Cooking time (min)	Na	K	P	Mg	Fe	Ca	Na/K	Ca/P
FCW	0	60.80±0.60 <sup>d</sup>	1109.30±0.81 <sup>c</sup>	259.25±1.08 <sup>a</sup>	140.60±0.52 <sup>f</sup>	10.33±0.29 <sup>b</sup>	340.50±0.51 <sup>b</sup>	0.05	1.31
	45	55.96±0.54 <sup>c</sup>	804.70±0.30 <sup>j</sup>	200.63±0.35 <sup>d</sup>	110.34±0.36 <sup>b</sup>	7.80±0.20 <sup>d</sup>	272.88±0.38 <sup>c</sup>	0.07	1.36
	60	52.30±0.70 <sup>f</sup>	716.20±0.75 <sup>k</sup>	163.69±0.31 <sup>k</sup>	90.75±0.18 <sup>g</sup>	5.63±0.15 <sup>h</sup>	240.23±0.68 <sup>e</sup>	0.07	1.46
	75	50.16±0.15 <sup>g</sup>	665.34±0.41 <sup>l</sup>	138.60±0.36 <sup>i</sup>	80.86±0.16 <sup>i</sup>	4.40±0.10 <sup>g</sup>	218.16±0.77 <sup>h</sup>	0.07	1.57
FCR	0	72.65±0.32 <sup>h</sup>	1592.90±6.38 <sup>a</sup>	250.48±0.67 <sup>b</sup>	140.56±0.20 <sup>f</sup>	12.56±0.28 <sup>a</sup>	359.32±0.89 <sup>a</sup>	0.05	1.43
	45	68.34±0.43 <sup>i</sup>	1150.22±0.23 <sup>d</sup>	196.57±0.50 <sup>c</sup>	106.32±1.14 <sup>c</sup>	9.26±0.14 <sup>c</sup>	300.69±0.27 <sup>k</sup>	0.06	1.52
	60	66.33±0.59 <sup>b</sup>	966.99±0.01 <sup>h</sup>	164.09±0.09 <sup>k</sup>	90.45±0.48 <sup>h</sup>	7.35±0.15 <sup>de</sup>	268.67±0.29 <sup>d</sup>	0.07	1.63
	75	64.22±0.23 <sup>c</sup>	852.50±0.50 <sup>i</sup>	145.14±0.15 <sup>h</sup>	81.12±0.13 <sup>i</sup>	6.30±0.20 <sup>th</sup>	233.60±0.40 <sup>f</sup>	0.07	1.6
FCB	0	75.27±0.72 <sup>a</sup>	1519.50±1.40 <sup>b</sup>	230.18±1.13 <sup>c</sup>	128.46±0.35 <sup>a</sup>	9.49±0.27 <sup>c</sup>	301.03±1.00 <sup>k</sup>	0.05	1.31
	45	71.90±0.26 <sup>h</sup>	1238.34±0.36 <sup>c</sup>	185.13±0.13 <sup>f</sup>	102.72±0.23 <sup>d</sup>	6.97±0.11 <sup>ef</sup>	223.84±0.96 <sup>g</sup>	0.06	1.21
	60	69.62±0.54 <sup>i</sup>	1100.92±0.08 <sup>f</sup>	150.33±0.15 <sup>g</sup>	91.97±0.24 <sup>g</sup>	5.86±0.04 <sup>h</sup>	203.81±0.19 <sup>j</sup>	0.06	1.35
	75	68.27±0.30 <sup>i</sup>	992.09±0.34 <sup>g</sup>	132.18±0.12 <sup>j</sup>	82.90±0.13 <sup>e</sup>	4.90±0.01 <sup>g</sup>	180.50±0.40 <sup>j</sup>	0.07	1.36

Mean  $\pm$  SD, n = 3; in columns, means marked with different letters indicate significant differences at the threshold ( $P < 0.05$ ). FCW (White Cultivar Flour), FCR (Red Cultivar Flour) and FCB (Black Cultivar Flour).

### 3.3. Effect of Cooking Time on Amino Acids Composition of *Phaseolus Lunatus* Bean Seeds

Nine amino acids, including 6 essential amino acids (lysine, phenylalanine, tryptophan, threonine, tyrosine and methionine) and three non-essential amino acids (arginine, cystine and glutamic acid), were determined after cooking *Phaseolus lunatus* bean seeds (Table 3). Amino acids in *P. lunatus* bean seeds did not decrease significantly ( $P < 0.05$ )

with cooking time. The most abundant essential amino acids are lysine, threonine, phenylalanine and tyrosine. Contents vary respectively from  $4.44 \pm 0.03$  -  $6.00 \pm 0.10\%$ ;  $6.38 \pm 0.03$  -  $9.80 \pm 0.10\%$ ;  $2.90 \pm 0.01$  -  $4.80 \pm 0.01\%$  and  $3.62 \pm 0.08$  -  $5.20 \pm 0.02\%$ . The most abundant non-essential amino acids are arginine and glutamic acid. The contents vary respectively from  $5.96 \pm 0.05$  -  $8.98 \pm 0.22\%$  and  $14.61 \pm 0.06$  -  $16.90 \pm 0.10\%$ . Seeds cooked for 45 minutes from the white cultivar showed low amino acid losses.

Table 3. Variation of aminoacids levels in *Phaseolus lunatus* bean seeds as a function of cooking time (mg/100gdrymatter).

Flours	Cooking time (min)	Cystine	Thréonine	Tryptophane	Tyrosine	Méthionine	Lysine	Phénylalanine	Arginine	Acide glutamique
FCW	0	0.55±0.01 <sup>fg</sup>	4.80±0.01 <sup>a</sup>	2.89±0.09 <sup>ef</sup>	5.20±0.02 <sup>c</sup>	0.58±0.10 <sup>g</sup>	5.41±0.12 <sup>fg</sup>	9.80±0.10 <sup>g</sup>	8.98±0.22 <sup>a</sup>	14.53±0.25 <sup>b</sup>
	45	0.50±0.01 <sup>fg</sup>	4.56±0.04 <sup>b</sup>	2.78±0.01 <sup>df</sup>	5.15±0.02 <sup>c</sup>	0.51±0.10 <sup>hi</sup>	5.32±0.02 <sup>fg</sup>	9.69±0.02 <sup>g</sup>	8.87±0.10 <sup>a</sup>	14.28±0.01 <sup>bf</sup>
	60	0.48±0.01 <sup>fg</sup>	4.37±0.02 <sup>c</sup>	2.72±0.02 <sup>d</sup>	5.11±0.01 <sup>c</sup>	0.46±0.10 <sup>ij</sup>	5.21±0.01 <sup>dg</sup>	9.61±0.04 <sup>bg</sup>	8.76±0.03 <sup>a</sup>	14.18±0.02 <sup>f</sup>
	75	0.46±0.01 <sup>g</sup>	4.27±0.02 <sup>c</sup>	2.66±0.01 <sup>d</sup>	5.07±0.02 <sup>c</sup>	0.44±0.10 <sup>j</sup>	5.11±0.01 <sup>de</sup>	9.50±0.10 <sup>b</sup>	8.68±0.07 <sup>a</sup>	14.07±0.06 <sup>f</sup>



Flours	Cooking time (min)	Cystine	Thréonine	Tryptophane	Tyrosine	Méthionine	Lysine	Phénylalanine	Arginine	Acide glutamique
FCR	0	1.07±0.01 <sup>a</sup>	3.52±0.02 <sup>c</sup>	3.64±0.04 <sup>b</sup>	4.32±0.07 <sup>a</sup>	1.37±0.20 <sup>a</sup>	5.00±0.10 <sup>c</sup>	8.36±0.03 <sup>a</sup>	6.20±0.20 <sup>b</sup>	16.90±0.10 <sup>a</sup>
	45	0.93±0.11 <sup>ac</sup>	3.30±0.02 <sup>f</sup>	3.57±0.06 <sup>bc</sup>	4.21±0.01 <sup>ad</sup>	1.24±0.10 <sup>b</sup>	4.63±0.05 <sup>h</sup>	8.14±0.05 <sup>c</sup>	6.10±0.10 <sup>b</sup>	16.70±0.04 <sup>ac</sup>
	60	0.82±0.09 <sup>cde</sup>	3.20±0.02 <sup>gh</sup>	3.52±0.02 <sup>bc</sup>	4.13±0.20 <sup>ad</sup>	1.17±0.10 <sup>f</sup>	4.52±0.03 <sup>h</sup>	8.00±0.10 <sup>ce</sup>	6.02±0.04 <sup>b</sup>	16.59±0.08 <sup>c</sup>
	75	0.77±0.02 <sup>de</sup>	3.13±0.01 <sup>gh</sup>	3.47±0.02 <sup>c</sup>	4.08±0.07 <sup>d</sup>	1.12±0.20 <sup>f</sup>	4.44±0.03 <sup>h</sup>	7.88±0.07 <sup>ef</sup>	5.96±0.05 <sup>b</sup>	16.53±0.03 <sup>c</sup>
FCB	0	0.86±0.05 <sup>cd</sup>	3.25±0.02 <sup>fg</sup>	3.09±0.01 <sup>a</sup>	4.03±0.15 <sup>d</sup>	0.90±0.03 <sup>c</sup>	6.00±0.10 <sup>a</sup>	7.72±0.02 <sup>f</sup>	7.30±0.10 <sup>c</sup>	15.44±0.03 <sup>d</sup>
	45	0.70±0.02 <sup>c</sup>	3.16±0.01 <sup>gh</sup>	2.93±0.03 <sup>c</sup>	3.80±0.10 <sup>b</sup>	0.1±0.02 <sup>d</sup>	5.71±0.03 <sup>b</sup>	7.53±0.05 <sup>h</sup>	7.13±0.15 <sup>c</sup>	15.31±0.01 <sup>de</sup>
	60	0.61±0.01 <sup>f</sup>	3.07±0.06 <sup>gh</sup>	2.87±0.06 <sup>ef</sup>	3.67±0.02 <sup>b</sup>	0.64±0.01 <sup>e</sup>	5.63±0.05 <sup>bc</sup>	7.45±0.05 <sup>h</sup>	7.05±0.12 <sup>c</sup>	15.24±0.04 <sup>de</sup>
	75	0.55±0.01 <sup>fg</sup>	2.90±0.01 <sup>d</sup>	2.87±0.03 <sup>ef</sup>	3.62±0.08 <sup>b</sup>	0.54±0.02 <sup>gh</sup>	5.50±0.10 <sup>cf</sup>	6.38±0.03 <sup>h</sup>	6.96±0.05 <sup>c</sup>	15.15±0.02 <sup>e</sup>

Mean ± SD, n = 3; in columns, means marked with different letters indicate significant differences at the threshold ( $P < 0.05$ ). FCW (White Cultivar Flour), FCR (Red Cultivar Flour) and FCB (Black Cultivar Flour).

### 3.4. Effect of Cooking Time on Functional Properties of Phaseolus Lunatus Bean Flours

#### 3.4.1. Water Absorption Capacity (WAC) and Water Solubility index (WSI)

The study showed a significant increase ( $P < 0.05$ ) in the water absorption capacity (WAC) and water solubility index (WSI) of seed flours from the three cultivars during cooking. The water absorption capacities of the cultivar flours (white, red and black) ranged from  $302.00 \pm 1.00$  -  $324.53 \pm 0.50$ ;  $246.53 \pm 0.50$  -  $266.43 \pm 0.51$  and  $230.00 \pm 1.00$  -  $257.53 \pm 1.00$  respectively. The water solubility index (WSI) of the

seed flours increased during cooking. The water solubility indices (WSI) of cultivar flours (white, red and black) range from  $40.50 \pm 0.50$  -  $43.10 \pm 0.27\%$ ;  $35.13 \pm 0.47$  -  $36.66 \pm 0.61\%$  and  $30.70 \pm 0.65$  -  $32.03 \pm 0.15\%$  respectively. Seed flours baked at 45 and 60 minutes have higher absorption capacities and water solubility indices (WSI) than seed flours baked at 75 minutes. All seed flours from the three cultivars have high water absorption capacities and water solubility indices (WSI) at 45 minutes cooking time. Flour from the white cultivar retained more water, with a high solubility index at 45 minutes of cooking, than flour from the red and black cultivars (Table 4).

Table 4. Evolution of water absorption capacity and solubility index of Phaseolus lunatus seed flours as a function of cooking time.

Flours	Cooking time (min)	Water absorption capacity (%)	water solubility index (%)
FCW	0	$302.00 \pm 1.00^b$	$33.10 \pm 0.43^e$
	45	$319.20 \pm 1.31^a$	$40.50 \pm 0.50^c$
	60	$324.53 \pm 0.50^c$	$45.36 \pm 0.32^b$
	75	$322.60 \pm 0.52^c$	$43.10 \pm 0.27^a$
FCR	0	$246.53 \pm 0.50^c$	$26.30 \pm 0.30^e$
	45	$262.43 \pm 0.51^b$	$35.13 \pm 0.47^i$
	60	$269.00 \pm 1.00^f$	$38.16 \pm 0.76^i$
	75	$266.43 \pm 0.51^f$	$36.66 \pm 0.61^d$
FCB	0	$230.00 \pm 1.00^d$	$22.30 \pm 0.26^f$
	45	$255.00 \pm 1.00^h$	$30.70 \pm 0.65^h$
	60	$260.63 \pm 1.52^h$	$34.80 \pm 0.80^h$
	75	$257.00 \pm 1.00^g$	$32.03 \pm 0.15^i$

Mean ± SD, n = 3; in columns, means marked with different letters indicate significant differences at the threshold ( $P < 0.05$ ). FCW (White Cultivar Flour), FCR (Red Cultivar Flour) and FCB (Black Cultivar Flour).

#### 3.4.2. Oil Absorption Capacity of Three Phaseolus Lunatus Cultivars Flours

The results show a significant increase (at the 5% threshold) in the oil absorption capacity of *P. lunatus* seed flours during cooking. The oil absorption capacities of uncooked *P. lunatus* seed flours are lower than those of cooked flours. The corn oil absorption capacity (CAH) of seed flours from *P. lunatus* cultivars ranged from  $190.36 \pm 0.40$  to  $228.93 \pm 0.81\%$  respectively. It is higher in the flour of the red cultivar, ranging from  $207.00 \pm 1.00$  -  $228.93 \pm 0.81\%$ . The sunflower oil absorption capacity of seed flours from *P. lunatus* cultivars ranged from  $210.30 \pm 0.30$  -  $245.36 \pm 0.40\%$  and was highest in the flour of the red cultivar. It range from  $220.86 \pm 0.80$  to  $245.36 \pm 0.40\%$ . The red oil absorption capacity of seed flours from *P. lunatus* cultivars ranged from  $199.50 \pm 0.50$  -  $240.03 \pm 0.45\%$ . Seed flour

from the red cultivar showed the highest red oil absorption capacity. It range from  $219.00 \pm 1.00$  -  $240.36 \pm 0.45\%$ . The dinor oil absorption capacity of seed flours from *P. lunatus* cultivars ranges from  $214.73 \pm 0.23$  -  $250.36 \pm 0.40\%$ . It was highest in the flour of the red cultivar, ranging from  $200.96 \pm 0.25$  -  $228.93 \pm 0.8\%$ . Raw cultivar flours have a significantly ( $P < 0.05$ ) lower CAH than cooked flours obtained at different cooking times. Cultivar flours have different CAHs depending on the oil studied (Table 5). At the 75 minutes seed cooking time, the oil absorption capacities of the flours are higher than at the 45 and 60 minutes times studied. The absorption of each oil is specific to a given flour. Flour from seeds of the red cultivar absorbs more dinor and sunflower oil, while flour from the white cultivar absorbs more red oil, and finally, corn oil is absorbed more by flour from the black cultivar.

**Table 5.** Oil absorption capacity of *Phaseolus lunatus* seed flours as a function of cooking time.

Flours	Cooking time (min)	Oil absorption Capacity (%)			
		Maize	Sunflower	Red	Dinor
FCW	0	200.00±1.00 <sup>i</sup>	210.30±0.30 <sup>g</sup>	219.00±1.00 <sup>h</sup>	214.73±0.23 <sup>d</sup>
	45	209.70±0.75 <sup>c</sup>	218.50±0.50 <sup>e</sup>	229.40±0.52 <sup>c</sup>	225.60±0.52 <sup>i</sup>
	60	215.30±1.08 <sup>g</sup>	224.03±0.55 <sup>h</sup>	235.06±0.40 <sup>c</sup>	233.46±1.40 <sup>h</sup>
	75	220.33±0.35 <sup>c</sup>	230.23±0.49 <sup>i</sup>	240.03±0.45 <sup>a</sup>	240.40±0.52 <sup>f</sup>
FCR	0	190.36±0.40 <sup>f</sup>	220.86±0.80 <sup>d</sup>	199.50±0.50 <sup>j</sup>	224.60±0.52 <sup>j</sup>
	45	200.96±0.25 <sup>i</sup>	231.00±1.00 <sup>i</sup>	209.53±0.50 <sup>i</sup>	237.00±1.00 <sup>g</sup>
	60	206.33±0.61 <sup>h</sup>	238.63±0.55 <sup>b</sup>	214.53±0.50 <sup>k</sup>	243.36±0.32 <sup>c</sup>
	75	213.26±0.30 <sup>d</sup>	245.36±0.40 <sup>a</sup>	221.13±0.32 <sup>g</sup>	250.36±0.40 <sup>a</sup>
FCB	0	207.00±1.00 <sup>h</sup>	213.26±0.64 <sup>f</sup>	215.53±0.50 <sup>k</sup>	220.36±1.00 <sup>c</sup>
	45	216.50±0.50 <sup>g</sup>	223.30±0.30 <sup>h</sup>	225.40±0.45 <sup>f</sup>	230.30±0.60 <sup>b</sup>
	60	223.10±0.36 <sup>b</sup>	229.56±0.51 <sup>i</sup>	231.36±0.40 <sup>d</sup>	235.36±0.32 <sup>g</sup>
	75	228.93±0.81 <sup>a</sup>	235.20±0.34 <sup>c</sup>	237.30±0.30 <sup>b</sup>	241.36±0.40 <sup>ef</sup>

Mean ± SD, n = 3; in columns, means marked with different letters indicate significant differences at the threshold ( $P < 0.05$ ). FCW (White Cultivar Flour), FCR (Red Cultivar Flour) and FCB (Black Cultivar Flour).

### 3.4.3. Changes in the Swelling of Seed Flours from Three *Phaseolus Lunatus* Cultivars as a Function of Cooking Time

The study showed that raw and cooked seed flours from all three *Phaseolus lunatus* cultivars swell with increasing incubation temperature. Swelling is low for all flours at incubation temperatures of 50-60°C, and increases from 70°C up to 90°C. Flours from seeds cooked at times 45, 60

and 75 from the three *P. lunatus* cultivars absorb  $16.50 \pm 0.45$  -  $18.53 \pm 0.20$  g water / g DM;  $18.56 \pm 0.51$ - $20.70 \pm 0.36$  g water / g DM and  $21.90 \pm 0.85$  -  $23.30 \pm 0.30$  g water / g DM respectively, while uncooked flour retains  $14.80 \pm 0.20$  - $15.53 \pm 0.30$  g water / g DM at 90°C. Cooked seed flours swell more than uncooked seed flours. At 90°C, flour from white cultivar seeds swells more than flour from red and black cultivars (Table 6).

**Table 6.** Swelling of *Phaseolus lunatus* seed flours as a function of cooking time.

Flours	Cookingtime (min)	Swelling (g water uptake/g DM)				
		50°C	60°C	70°C	80°C	90°C
FCW	0	2.33±0.25 <sup>e</sup>	5.33±0.41 <sup>cf</sup>	8.43±0.51 <sup>e</sup>	12.43±0.51 <sup>de</sup>	15.43±0.51 <sup>fg</sup>
	45	4.66±0.32 <sup>d</sup>	8.63±0.15 <sup>c</sup>	10.73±0.64 <sup>d</sup>	15.56±0.51 <sup>f</sup>	18.70±0.26 <sup>de</sup>
	60	6.90±0.79 <sup>b</sup>	10.93±0.40 <sup>d</sup>	13.40±0.36 <sup>f</sup>	17.93±0.11 <sup>b</sup>	20.70±0.36 <sup>c</sup>
	75	10.50±0.26 <sup>c</sup>	12.00±0.20 <sup>bd</sup>	15.70±0.81 <sup>a</sup>	20.46±0.41 <sup>a</sup>	23.30±0.30 <sup>a</sup>
FCR	0	2.26±0.30 <sup>e</sup>	4.43±0.40 <sup>c</sup>	6.53±0.50 <sup>b</sup>	11.60±0.52 <sup>ce</sup>	15.53±0.30 <sup>fg</sup>
	45	5.36±0.20 <sup>d</sup>	6.46±0.37 <sup>f</sup>	8.10±0.36 <sup>bc</sup>	13.60±0.52 <sup>d</sup>	18.83±0.20 <sup>c</sup>
	60	7.80±0.26 <sup>ab</sup>	9.03±0.25 <sup>c</sup>	11.26±0.55 <sup>cd</sup>	14.96±0.35 <sup>f</sup>	19.20±0.43 <sup>d</sup>
	75	10.76±0.25 <sup>c</sup>	12.43±0.37 <sup>ab</sup>	12.83±0.15 <sup>cf</sup>	16.06±0.25 <sup>f</sup>	2.10±0.36 <sup>ab</sup>
FCB	0	3.10±0.30 <sup>e</sup>	5.86±0.70 <sup>f</sup>	8.96±0.35 <sup>c</sup>	10.60±0.52 <sup>c</sup>	14.80±0.20 <sup>e</sup>
	45	5.30±0.30 <sup>d</sup>	8.16±0.15 <sup>c</sup>	11.06±0.80 <sup>d</sup>	12.83±0.76 <sup>de</sup>	16.50±0.45 <sup>f</sup>
	60	8.63±0.30 <sup>a</sup>	10.63±0.65 <sup>d</sup>	12.76±0.68 <sup>cf</sup>	15.16±0.28 <sup>f</sup>	18.56±0.51 <sup>de</sup>
	75	11.53±0.50 <sup>c</sup>	13.83±0.97 <sup>a</sup>	13.86±0.73 <sup>f</sup>	18.40±0.36 <sup>b</sup>	21.90±0.85 <sup>bc</sup>

Mean ± SD, n = 3; in columns, means marked with different letters indicate significant differences at the threshold ( $P < 0.05$ ). FCW (White Cultivar Flour), FCR (Red Cultivar Flour) and FCB (Black Cultivar Flour).

### 3.4.4. Evolution of the Solubility of Seed Flours of the Three *Phaseolus Lunatus* Cultivars as a Function of Cooking Time

The results of the flour solubility analysis show that the solubility of cooked *Phaseolus lunatus* seed flours is higher than that of uncooked flours. There was a significant difference ( $P < 0.05$ ) whatever the cooking temperature and time (Table 7). The solubility of flours obtained at 45, 60 and

75 minutes baked is higher at 90°C than at 50°C, 60°C, 70°C and 80°C. They range from  $12.83 \pm 0.20$  to  $15.56 \pm 0.49\%$ ;  $15.70 \pm 0.36$  to  $17.83 \pm 0.15\%$  and  $17.30 \pm 0.30$  to  $19.90 \pm 0.85\%$ . The solubility of uncooked flours ranged from  $10.53 \pm 0.30$  to  $13.46 \pm 0.41\%$ . Uncooked seed flour is less soluble than cooked seed flour. Solubility increases with cooking time. Black cultivar flour is more soluble than red and white cultivar flour at 90°C.

**Table 7.** Evolution of *Phaseolus lunatus* seed flours solubility as a function of cooking time.

Flours	Cooking time (min)	Solubility (%)				
		50°C	60°C	70°C	80°C	90°C
FCW	0	1.80±0.20 <sup>c</sup>	3.33±0.41 <sup>g</sup>	5.23±0.30 <sup>b</sup>	8.16±0.15 <sup>h</sup>	11.10±0.17 <sup>b</sup>
	45	3.80±0.10 <sup>d</sup>	6.63±0.15 <sup>de</sup>	7.30±0.30 <sup>e</sup>	10.90±0.85 <sup>ef</sup>	13.36±0.32 <sup>e</sup>
	60	5.90±0.79 <sup>b</sup>	8.26±0.25 <sup>h</sup>	9.40±0.36 <sup>d</sup>	12.60±0.52 <sup>bc</sup>	15.70±0.36 <sup>d</sup>
	75	8.50±0.26 <sup>ac</sup>	9.50±0.30 <sup>bc</sup>	10.86±0.55 <sup>c</sup>	14.46±0.41 <sup>b</sup>	17.30±0.30 <sup>c</sup>
FCR	0	1.90±0.10 <sup>c</sup>	3.63±0.15 <sup>fg</sup>	4.53±0.50 <sup>b</sup>	7.03±0.25 <sup>h</sup>	10.53±0.30 <sup>b</sup>
	45	4.33±0.15 <sup>d</sup>	5.90±0.10 <sup>c</sup>	6.93±0.11 <sup>c</sup>	9.40±0.40 <sup>g</sup>	12.83±0.20 <sup>e</sup>
	60	6.00±0.20 <sup>b</sup>	8.76±0.25 <sup>ch</sup>	9.63±0.55 <sup>d</sup>	12.30±0.30 <sup>cd</sup>	15.86±0.15 <sup>d</sup>
	75	7.76±0.25 <sup>c</sup>	10.43±0.37 <sup>a</sup>	11.83±0.15 <sup>ac</sup>	14.03±0.25 <sup>b</sup>	17.76±0.25 <sup>c</sup>
FCB	0	1.96±0.15 <sup>c</sup>	4.4±0.26 <sup>f</sup>	6.30±0.30 <sup>e</sup>	9.80±0.20 <sup>fg</sup>	13.46±0.41 <sup>e</sup>
	45	4.53±0.32 <sup>d</sup>	7.16±0.15 <sup>d</sup>	8.86±0.15 <sup>e</sup>	11.16±0.28 <sup>de</sup>	15.56±0.49 <sup>d</sup>
	60	7.63±0.30 <sup>c</sup>	8.30±0.30 <sup>h</sup>	11.10±0.17 <sup>c</sup>	13.73±0.25 <sup>bi</sup>	17.83±0.15 <sup>c</sup>
	75	9.26±0.30 <sup>a</sup>	9.83±0.20 <sup>ab</sup>	12.53±0.20 <sup>a</sup>	15.73±0.25 <sup>a</sup>	19.90±0.85 <sup>a</sup>

Mean ± SD, n = 3; in columns, means marked with different letters indicate significant differences at the threshold ( $P < 0.05$ ). FCW (White Cultivar Flour), FCR (Red Cultivar Flour) and FCB (Black Cultivar Flour).

## 4. Discussion

The present study focused to investigate effect of cooking time on some biochemical parameters and functional properties of *Phaseolus lunatus* (L.) bean seeds. Firstly, some biochemical parameters were carried out.

Concerning water content of cooked *Phaseolus lunatus* (L.) bean seeds, it increased with cooking time. This is due to the nutrient dilution effect caused by the absorption of water by the seeds during cooking [18].

Levels of total and reducing sugar contents of cooked *P. lunatus* (L.) bean seeds decreased with increasing cooking time. The decrease in total and reducing sugars may be due to simultaneous hydrolysis and gelatinization of the seeds during cooking. Indeed, under the action of heat, starch granules well and rupture to release their contents, composed mainly of amylose and amylopectin [19]. These molecules are then hydrolyzed in to total and reducing sugars, which are then dispersed in the cooking water, resulting in their depletion. The decrease in sugars during seed cooking was reported by [20], who observed a drop in reducing sugars after cooking chickpea (*Cicer arietinum*) seeds. The total and reducing sugar contents of *P. lunatus* bean flours are higher than those observed by [21] in soybeans. They obtained total sugar contents of between 0.05 and 0.4% in uncooked and cooked soybean flours respectively, and also reducing sugar contents of between 0.03 and 0.04% of solids.

The drop in protein content during cooking of *Phaseolus lunatus* (L.) bean seeds is only significant after a cooking time of 45 min. This result can be explained by the effective denaturation of proteins after 45 min of cooking, i.e. a break in peptide bonds and certainly in protein disulphide bridges [22]. These results are consistent with those of [23], who reported that the crude protein content of cooked millet is lower than that of uncooked millet. According to [24], the protein needs of infants are estimated at 9g/day. The protein content of *P. lunatus* bean flours ranged from 19.03 to 25.06%, which may partially meet the protein requirements of infants.

The decrease of lipid content during cooking of *Phaseolus*

*lunatus* (L.) bean seeds may be due to the leaching of lipids into the cooking water [25]. This results are consistent with those of [26], who observed fat loss in cooked *Mucuna* spp seeds. [27] observed the same similarities in their work on *Vigna sesquipedalis* beans.

Ash content decreases in *Phaseolus lunatus* (L.) bean seeds during cooking. This could be explained by the leaching of minerals in boiling water. Similar results have also been reported for soybean (*glycine maximum*) and lima bean (*P. lunatus*), when samples were subjected to the autoclave sterilization process [28].

Crude fiber contents increase in *Phaseolus lunatus* bean seeds during cooking. This may be explained by the formation of protein-fiber complexes following the chemical modification induced by cooking of lentils (*lens culinaris*) [29]. Also, this increase in fiber in cooked *P. lunatus* bean seeds is similar to the work of [30, 31], who showed that cooking increases soluble fiber content and decreases insoluble fiber content. Consumption of *P. lunatus* seeds may help reduce the risk of hypertension, constipation, diabetes, colon cancer and breast cancer.

The carbohydrate content of *Phaseolus lunatus* bean seeds increases during cooking. This is due to the mathematical difference used to determine carbohydrate levels. This carbohydrate content of *P. lunatus* bean seeds may make this bean an energy food that can contribute to food security in developing countries [32], particularly in Côte d'Ivoire.

The energy value of *Phaseolus lunatus* bean seeds increases with cooking time as carbohydrates. This increase is similar to that observed in *Dioscorea alata* raw and cooked for 90 min (357.65 and 370.01Kcal/100g dry matter respectively) [33]. *Phaseolus lunatus* bean seeds could be used in part as an energy food in porridges for infants and children whose energy requirements range from 547 to 1092 kcal/day [34].

*Phaseolus lunatus* bean seeds cooked at 45 as cooking time contain most minerals. In general, minerals decreased significantly ( $P < 0.05$ ) with cooking time. The loss of minerals is due to the degradation of anti-nutritional factors such as phytate, which traps 60-80% of these minerals in seeds, compared with 20-34% in fruits and tubers [35], with

iron being trapped by tannin [36]. The findings [37, 38] have shown that the denaturation of anti-nutritional factors by heat during cooking of *P. lunatus* bean seeds will release minerals into their matrices, which will then diffuse into the cooking water [39], resulting in their reduction. The Na/K and Ca/P ratios of *P. lunatus* bean seeds are less than or greater than 1. The Na/K ratio is very important for the body, as sodium and potassium regulate high blood pressure and muscle contraction. A food product is a good source of Ca and Fe if the Ca/P ratio is greater than 1 [40]. The iron content of *P. lunatus* bean seeds can be recommended in the human diet because, according to [41], the recommended level for human consumption is 1.37mg/day (men) and 2.94mg/day (women). *P. lunatus* bean seeds can be recommended as a dietary supplement.

In general, cooked *Phaseolus lunatus* seeds show low aminoacid losses. Specifically, seeds of white cultivar cooked at 45 min showed low aminoacid losses compared with those from the red and black cultivars. Essential and non-essential amino acids decreased non-significantly ( $P > 0.05$ ) during cooking. The rate of reduction of essential aminoacids (Lysine, phenylalanine and tryptophan) in *P. lunatus* bean seeds during cooking is similar to the work of [42] in lentil seeds (*lens culinaris*). The lysine content of raw and cooked *P. lunatus* bean flours ranging from 4.44 to 6.00 mg/100g dry matter is lower than that of raw and cooked lentil (*lens culinaris*) seeds (6.9 to 7.00mg/100g dry matter [42]). Non-essential aminoacids (methionine and arginine) were slightly reduced during cooking of *P. lunatus* bean seeds. Our results are similar to those of [43] who showed the reduction of sulfur aminoacids during cooking of faba bean seeds. The essential amino-acid/total amino acid ratios of uncooked and 45 minute cooked seeds of the three *P. lunatus* cultivars ranged from 0.51 to 0.54 and 0.46 to 0.50%. Therefore, uncooked and 45 minute cooked seeds can be incorporated into the following diets.

Secondly, functional properties of *Phaseolus lunatus* (L.) bean seeds were carried out.

Flour from the white cultivar retains more water, with a high solubility index during cooking. The EAC values of uncooked and cooked *Phaseolus lunatus* seeds flours (230-322.60%) are within the range of the EACs of raw and pre cooked taro flours (247.5% and 562.5%) respectively [44], but higher than those of uncooked and cooked rice (225% and 250%) [45]. The high FAC values of cooked *Phaseolus lunatus* seed flours are thought to be linked to starch gelatinization, which increases water-binding capacity [46]. The high CAE values of *Phaseolus lunatus* seed flours suggest that they could be used in soup formulations for easier digestion [47]. [48] has shown that EAC is an important property of flours used in pastry-making, since it enables pastry-makers to add plenty of water to the dough while improving handling and maintaining freshness in bread. The high CAE values of *Phaseolus lunatus* bean seed flours suggest that they could be useful in soup formulations for easier digestion [47]. [48] showed that EAC is an im of the starch grains contained in the flours. The water solubility index (WSI) reflects the extent of starch degradation

[49]. The high WSI of uncooked *Phaseolus lunatus* bean flour may be due to gelatinization of the starch grains, which facilitates starch solubility [50]. The high ISE percentages of cooked *P. lunatus* bean flours show that cooking has had an effect on starch degradation. The ISE levels of *P. lunatus* bean flours (22 - 43.10%) are higher than those of raw taro flours (18% to 27%) [49] and those of raw and cooked yam (*Dioscorea spp*) flours (9.26% and 15.31%) respectively [51]. The oil absorption capacity of flours increases as the seeds are cooked. Oil absorption is specific to a given flour. Flour from seeds of the red cultivar absorbs more dinor and sunflower oil, while flour from the white cultivar absorbs more red oil, and finally, corn oil is absorbed more by flour from the black cultivar. Oil absorption capacity (OAC) is an important property in the formulation of a food product, as oil improves flavor and gives a smooth texture to the food [52, 53].

According to [54], CAH is influenced by proteins, but is mainly due to the availability of lipophilic groups. The ability of proteins to retain oil is an interesting property, as it enables good flavour retention during food processing, thus improving palatability [55]. The absorption capacities of maize, sunflower, red and dinor oils obtained respectively 190.36-228.93%; 210.30-245.36%; 199.50-240.03% and 214.13-250.36% are lower than those reported by [56], who observed an increase in the CAH of jackfruit flours from 280% to 310%. The CAH of *Phaseolus lunatus* bean flour is higher than that of yam flour (190%) [57]. Cooked seed flours swell more than uncooked seed flours. At 90°C, seeds flours from the white cultivar swell more than flours from other cultivars. The increase in flour swelling following heating is thought to be due to changes in the physical state of the starch granules in the flours during hydrothermal treatment.

In fact, under the effect of heat, the granules weaken, allowing water to penetrate them. During swelling, the amylose comes out of the granule and the amylopectin is strongly hydrated. As temperature increases, granules tend to rupture, collapse and fragment, releasing polymer molecules and aggregates [58]. The swelling power (SP) of uncooked and cooked flours varies from 10.53 to 19.90% at 90°C. Cooking significantly ( $p < 0.05$ ) increased the PG of the flours. The high PG of uncooked *Phaseolus lunatus* bean flours is due to the fact that proteins and lipids, by forming complexes with the starch granules in the flours, prevent the penetration of water into the granules, thus limiting the PG. This view is confirmed by [59], who reported that proteins and lipids limited the PG of rice granules during cooking. The PG of *P. lunatus* bean flours is higher than that of cassava flour (2.16%) [60] and that of flour from different rice varieties (10.76 to 12.97% at 90°C) [61]. PG is a good indication of water absorption by the granules during heating [62]. The solubility of the different *P. lunatus* bean seed flours increase with cooking time. Flour from black seeds is more soluble than flour from red and white cultivars at 90°C. The solubility values of cooked *P. lunatus* seed flours are higher than those of uncooked breadfruit flour. This result can be explained by the fact that cooked *P. lunatus* bean flours have a high PG. Granules with a high PG burst to release their contents into the reaction medium. The solubility



of *Phaseolus lunatus* seed flour is higher than that of cassava flour (7.57%-10.46%) [63]. Cooked *P. lunatus* bean flours are digestible and therefore suitable for infant food formulations.

## 5. Conclusion

A study of the effect of cooking time on the biochemical parameters and functional properties of *Phaseolus lunatus* seeds in water at 100°C for 45, 60 and 75 minutes showed a decrease in nutritional factors except for carbohydrates, energy value and fiber, which increased respectively from 64.16 to 70.39%, 300.79 to 374.28 Kcal/100 g and 4.62 to 6.05% dry matter. The energy value is highest with black seeds cooked for 45 minutes, whereas 60 minutes cooking of the same seeds and white yields high carbohydrate and fiber values. Nutritional factors such as minerals and amino acids are reduced during cooking. Water absorption, oil absorption, swelling capacity and solubility improved considerably with different cooking times. In view of the results, the best cooking time for *P. lunatus* seeds would be 45 min for good nutritional quality. Functional properties also show that cooked *P. lunatus* seed flours are suitable for the preparation of baby food porridges, pastries and cakes.

## References

- [1] Amini. R., Pezhgan. h. and Mohammadinasab. A. D. (2013). Effect of weeds competition on some growth parameters of red, white and pinto bean (*Phaseolus vulgaris* L.). *Journal of Biodiversity and Environmental Sciences (JBES)*. 3:86-93.
- [2] Corte's AJ, Monserrate FA, Ram1'-rez-Villegas J, Madrin- a'n S, Blair MW. (2013). Drought Tolerance in Wild Plant Populations: The Case of Common Beans (*Phaseolus vulgaris* L.). *PLoS ONE* 8 (4): e62898. doi: 10.1371/journal.pone.0062898.
- [3] Sparvoli F, Laureati M, Pilu R, Pagliarini E, Toschi 1, Giuberti G, Fortunati P, Daminati MG, Cominelli E and Bollini R. (2016). Exploitation of Common Bean Flours with Low Antinutrient Content for Making Nutritionally Enhanced Biscuits. *Front. Plant Sci.* 7: 928.
- [4] Kizito Iheanacho. M. E. (2010). Comparative studies of the nutritional composition of soybean (*Glycine max*) and Lima bean (*Phaseolus lunatus*). *Scientia Africana*, 9: 29-35.
- [5] Ezeagu E. I. & Ibegbu M. D. (2010). Biochemical composition and nutritional potential of ukpa: a variety of tropical Lima beans (*Phaseolus lunatus*) from Nigeria. *Polish Journal Food Nutrition Sciences*, 60 (3): 231-235.
- [6] Gardner C. D. I., Chatterjee C., Rigby A., Spiller G. & Farquhar J. W. (2005). Effects of a plant- based diet on plasma lipids in hypercholesterolemic Adults. *Annals International Med*, 142: 725-1155.
- [7] Andriamamonjy N. (2000). Valeur nutritionnelle des graines sèches de 7 variétés de haricot et de 2 variétés d'Ambérie (Mémoire de DEA de Biochimie appliquée aux sciences de l'alimentation et à la nutrition). Faculté des sciences: Université d'Antananarivo, 53p.
- [8] Tchumou. (2017): Ethnobotanical survey and physicochemical characterization of bean seeds, *Phaseolus lunatus* (Fabaceae) consumed in the South and East of Côte d'Ivoire according to maturity level and cooking time.
- [9] Yellavila, S. B., J. K. Agbenorhevi, J. Y. Asibuo and G. O. Sampson. (2015). Proximate composition, minerals content and functional properties of five lima bean accessions. *J. Food Secur.* 3: 69-74.
- [10] AOAC. (1990). Official methods of analysis of the Association of Official Analytical Chemists, 15th ed, Washington DC, 1230p.
- [11] Bernfeld. (1955). Amylase  $\beta$  and  $\alpha$ , In: method in enzymology 1, Colowick S. P. and Kaplan N. O., Academic Press, pp149-154.
- [12] FAO/INFOODS. (2015). FAO/INFOODS Guidelines for verifying food composition data before publication of a user table/database-Version 1.0. FAO, Rome.
- [13] Fatima Ismail., Farah N., Talpur. & Memon A. N. (2013). Determination of Water Soluble Vitamin in Fruits and Vegetables Marketed in Sindh Pakistan. *Pakistan Journal Nutrition*, 12: 197-199.
- [14] Oshodi A. A. (1992). Proximate composition, nutritionally valuable minerals and functional properties of *Adenopus breviflorus* benth seed flour and protein concentrate. *Food*.
- [15] Phillips R. D., Chinnan M. S., Branch A. L., Miller J. & Mcwatters K. H. (1988). Effects of pre-treatment on functional and nutritional properties of cowpea meal. *Journal of Food Sciences*, 53: 805-809.
- [16] Eke O. S. & Akobundu E. N. T. (1993). Functional properties of African yam bean (*Sphenostylis stenocarpa*) seed flour as affected by processing. *Food Chemical*, 48: 337-340.
- [17] Adebooye O. C. & Singh V. (2008). Physico-chemical properties of the flours and starches of two cowpea varieties (*Vigna unguiculata* L.) Walp). *Innovative Food Science and Emerging Technologies*, 9: 92-100.
- [18] Rehman A. R. & I. A. Khalil. (1988). Retention of selected nutrients in cooked kidney beans (*Phaseolus vulgaris* L.). *Journal of Agriculture of Food Chemistry*, 32: 342-346.
- [19] Bjorck I., ASP N. G., Birkhe D. & Lunquist I. (1984). Effects of processing on availability of starch digestion in vitro and in vivo. Extrusion cooking of wheat flours and starch. *Journal of Cereal Science*, 2: 91-103.
- [20] Alajaji S. A. & El-Adawy T. A. (2006). Nutritional composition of chickpea (*Cicer arietinum* L.) as affected by microwave cooking and other traditional cooking methods. *Journal of Food Composition and Analysis*, 19: 806-812.
- [21] Gupta V. & Nagar R. (2008). Physico-chemical and acceptability of rabadi (a fermented soya flour product) as affected by cooking and fermentation time. *International Journal of Food Science and Technology*, 43: 939-943.
- [22] Abeke F. O., Ogundipe S. O., Sokoni A. A., Dafwang I. I., Adeyinka I. A., Oni O. O. & Abeke A. (2007). Growth and subsequent egg production performance of shikabrown pullets fed graded levels of cooked *lablab purpureus* beans. *Pakistan Journal of Biological Sciences*, 10: 1051-1061.

- [23] Ocheme O. B., Oludamilola O. O. & Gladys M. E. (2010). Effect of lime soaking and cooking (nixtamalization) on the proximate functional and some anti-nutritional properties of millet flour. *Assumption University Journal of Technology*, 14: 131- 138.
- [24] FAO / WHO (Food & Agriculture Organization & the World Health Organization). (2007). Protein and amino acid requirements in human nutrition. Report of a Joint WHO / FAO / UNU Expert Technical Report Series 935. Cholé-Doc N°111.
- [25] Okaka J. C., Akobundu E. N. T. & Okaka A. N. C. (1992). Human Nutrition – An Intergrated Approach. Obio Press Ltd., Enugu, Pp. 182-220.
- [26] Vadivel V. & Pugalenth M. (2009). Effect of soaking in sodium bicarbonate solution followed by autoclaving on the nutritional and antinutritional properties of velvet bean seeds. *Journal Food Process Preservation*, 33: 60-73.
- [27] Nzewi D. & Egbuonu A. C. C. (2011). Effect of boiling and roasting on the proximate properties of asparagus bean (*Vigna Sesquipedalis*). *African Journal of Biotechnology*, 10: 11239-11244.
- [28] Aletor V. A. (1993). Allelochemicals in plant foods and feeding Stuffs. Part I. Nutritional, Biochemical and Physiopathological aspects in animal production. *Veternary and Human Toxicology*, 35: 57-67.
- [29] Bressani T. (1993). Grain quality of common beans. *Food Review International* 9: 237–297.
- [30] Slavin J. L. (1987). Dietary fiber, and body weight. *Journal of American Dietetic Association*, 87: 1164-1168.
- [31] Lintas C & Cappelloni M. (1998). Dietary fiber, resistant starch and in vitro starch digestibility of cereal meals. *Food Science and Nutrition*, 42: 117-124.
- [32] FAO. (2001). La nutrition dans les pays en voie de développement. FAO Ed, Genève, 515 pp.
- [33] Ezeocha V. C. & Ojmelukwe P. C. (2012). The impact of cooking on the proximate composition and anti-nutritional factors of water yam (*Dioscorea alata*). *Journal of Stored Products and Postharvest Research*, 3: 172 - 176.
- [34] Butte N. F. (1996). Energy requirements of infants. *European Journal of Clinical Nutrition* 50: 24-36.
- [35] Tran G. & Skiba F. (2005). Variabilité inter et intra matière première de la teneur en phosphore total et phytique et de l'activité phytasique. *INRA Productions Animales*, 18: 159- 168.
- [36] Brune M., Hallberg L. & Skanberg A. B. (1991). Determination of iron-binding phenolic groups in foods. *Journal of Food Science*, 56: 131-137.
- [37] Alonso R., Rubio L. A., Muzquiz M. & Marzo F. (2001). The effect of extrusion cooking on mineral bioavailability in pea and kidney bean seed meals. *Animal Feed Sciences and Technology*, 94: 1-13.
- [38] Anigo I. A., Ameh D. A., Ibrahim S. & Danbauchi S. S. (2009). Nutrient composition of commonly used complimentary foods in north western Nigeria. *African Journal of Biotechnology* 8: 4211- 4216.
- [39] Ewald C., Fjellkner-Modig S., Johansson K., Sjöholm I. & Akesson B. (1999). Effect of processing on major flavonoids in processed onions, green beans, and peas. *Food Chemistry*, 64: 231-235.
- [40] Nieman D. C., Butterworth. & Nieman C. N. (1992). Nutrition, WmC. Brown publishers. Dubugue, USA, 237-312pp.
- [41] Siddhuraju P. & Becker K. (2001). Effect of various domestic processing methods on antinutrients and in vitro-protein and starch digestibility of two indigenous varieties of Indian pulses, *Mucuna pruriens* var *utilis*. *Journal Agriculture of Food Chemistry*, (49): 3058-3067.
- [42] Hefnawy. T. H. (2011). Effect of processing methods on nutritional composition and antinutritional factors in lentils (*Lens culinaris*), 56: 57-61.
- [43] Khalil A. H. & Mansour E. H. (1995). The effect of cooking, autoclaving and germination on the nutritional quality of faba beans. *Food Chemistry*, 54: 177–182.
- [44] Fagbemi T. N. & Olaofe O. (2000). The chemical composition and functional properties of raw and precooked Taro (*Cococasia esculenta*) flour. *Journal of Biology Physical and Science*, 1: 98-103.
- [45] Abulude F. O. (2004). Effects of processing on nutritional composition, phytate and functional properties of rice (*Oryza sativa*) flour. *Nigerian Food Journal*, 22: 97-104.
- [46] Dengate H. N. (1984). Swelling, pasting and gellin of wheat starch, in *Advances in Cereal and Technology* (eds). By Pomeranz Y. AACC Eagan Press, St Paul, MN, 2: 49-84.
- [47] Olaofe O., Adeyemi F. O. & Adediran G. O. (1994). Amino acid and mineral and functional properties of some Oilseeds. *Journal of Agricultural and Food Chemistry*, 42: 878-881.
- [48] Wolf W. J. (1970). Soybean proteins: their functional, chemical and physical properties. *Journal of Agriculture and Food Chemistry*, 18: 965-969.
- [49] Mbofung C. M. F., Aboubakar N. Y., Njintang A., Bouba A. & Balaam F. (2006). Physicochemical and functional properties of six varieties of taro (*Colocasia esculenta* L.) flours. *Journal of Food Technology*, 4: 135-142.
- [50] Altan A., McCarthy K. L. & Maskan M. (2009). Effect of extrusion coking on functional properties and in vitro starch digestibility of barley-based extrudates from fruit and vegetable by-products. *Journal of Food Science*, 74: 77- 86.
- [51] Hsu C. L., Chen W., Weng Y. M. & Tseng C. Y. (2003). Chemical composition, physical properties, and antioxidant activities of yam flours as affected by different drying methods. *Food Chemistry*, 83: 85-92.
- [52] Aremu M. O., Olonisakin A; Atolaye B. O. & Ogbu C. F. (2006). Some nutritional and functional studies of *Prosopis africana*. *Electronic Journal of Environmental, Agricultural and Food Chemistry*, 5: 1640-1648.
- [53] Ubbor S. C. & Akobundu E. N. T. (2009). Quality Characteristics of Cookies from Composite Flours of Watermelon Seed, Cassava and Wheat. *Pakistan Journal of Nutrition*, 8: 1097-1102.
- [54] El-Adawy T. A. (2000). Functional properties and nutritional quality of acetylated succinylated mung bean protein isolate. *Food Chemistry*, 70: 83-91.
- [55] Moure A., Sineiro J., Dominguez H. & Parajo J. C. (2006). Fonctionnalité of oil seed protein product: A review. *Food Resaerch International*, 38: 945-963.

- [56] Odoemelam S. A. (2005). Functional properties of raw and heat processed jackfruit (*Artocarpus heterophyllus*) flour. *Pakistan Journal of Nutrition*, 4: 366-370.
- [57] Ukpabi U. J. (2010). Farmstead bread making potential of lesser yam (*Dioscorea esculenta*) flour in Nigeria. *Australian Journal of Crop Science*, 4: 68-73.
- [58] James P. C. (1980). Pulp and Paper Chemistry and Chemical Technology, John Wiley, New York, 3ème edition, 3: 1475-1508.
- [59] Ogawa Y., Glenn G. M., Orts W. J. & Wood D. F. (2003). Histological structures of cooked rice grain. *Journal of Agriculture and Food Chemistry*, 51: 7019-7023.
- [60] Okezie B. O. & Bello A. B. (1988). Physicochemical and functional properties of winged bean flours and isolate compared with soy isolate. *Journal of Food Science*, 53: 445 - 450.
- [61] Yu S., Ying M., Menager L. & Da-Wen S. (2012). Physicochemical Properties of Starch and Flour from Different Rice Cultivars. *Food Bioprocess Technology*, 5: 626-637.
- [62] Loos P. J., Hood L. F. & Graham H. D. (1981). Isolation and characterization of starch from breadfruit. *Cereal Chemistry*, 58: 282-286.
- [63] Baafi E. & Safo-Kantanka O. (2007). Effect of genotype, age and location on cassava flour yield and quality. *Journal of Plant Science*, 2: 607-612.